

Effect of salt stress on activity of superoxide dismutase (SOD) in *Ulmus pumila* L.

SONG Fu-nan, YANG Chuan-ping*, LIU Xue-mei, LI Gong-bin

Northeast Forestry University, Harbin 150040, P. R. China

Abstracts: The injury tolerance of cell plasma membrane and the correlative enzymes activities of plasma-membrane protection system in the *Ulmus pumila* leaves treated by nine concentrations (0.3%, 0.6%, 0.9%, 1.2%, 1.5%, 1.8%, 2.1%, 2.4%, 3.0%) of Na_2CO_3 and NaHCO_3 mixtures were studied in a greenhouse of Northeast Forestry University, Harbin, China. The rate of electrolyte leakage (REL) and SOD (Superoxide dismutase) activity in leaves of different samples were determined. Results showed that the REL in leaves of *U. pumila* presented a slowly increasing trend at the salt concentrations less than 1.5%, which indicated that cell plasma membrane of *U. pumila* leaves had rather strong resistance to the injury of salt ion, and had a significant increase at the salt concentrations more than 1.5%. The SOD activities in leaves of *U. pumila* presented an increased trend at salt concentrations less than 1.5%, the growth of seedlings did not decline, and tress and leaves had no symptom of injury, while the salt concentrations exceeded 1.5%, SOD activities sharply decreased and REL increased promptly.

Keywords: *Ulmus pumila*; Salt stress; Rate of electrolyte leakage (REL); SOD

CLC number: Q945.78

Document code: A

Article ID: 1007-662X(2006)01-0013-04

Introduction

Salt tolerance of plants mainly depended on the stability of cell membrane system which can still keep the integrality under salt stress to maintain the selective absorption for ion and other physiological function of cell (Greenway & Munne 1980). Salinity induced a wide range of responses in plants, and severe salt stress may cause oxidative damages, ion toxicity, nutritious imbalance, and plant death (Xiong *et al.* 2002). During normal metabolism, plants generated reactive oxygen species (ROS), including superoxide radical (O_2^-), hydrogen peroxide (H_2O_2), hydroxyl radical (HO^\cdot), and singlet oxygen (O_2^1) (Zhang *et al.* 2005), and the generation and elimination of ROS kept a dynamic balance at all times. When plants were stressed by bad environment, this balance was broken and ROS was overproduced in plants. Meantime, the biosynthesis of some antioxidant enzymes (SOD, POD, PPO, and MDA, CAT, and so on) in plants was induced by superabundant ROS, which can repair the DNA injury in time to maintain the normal growth of plants. Many microorganisms and plants also had the function inducing antioxidant defense system under environmental stresses (Shi *et al.* 2004; Ireneusz *et al.* 2003). ROS induced by salt stress played important roles in adaptive responses at lower concentrations, while it caused damages to macromolecules, such as DNA, proteins, membrane lipids, at higher concentrations (Wang *et al.* 2004; Mittler 2002; Breusegem *et al.* 2001). Injury caused by ROS is known as oxidative stress, which is the major cause of damage in plants exposed to different stressors (Bowler *et al.* 1992). Therefore much attention has been paid in recent years to the capacity that balanced the generation and elimination of ROS

in plant cells under environmental stresses (Mittler 2002). Plants have well-developed defense systems against ROS, but the mechanisms are not well understood (Alscher *et al.* 2002). Superoxide dismutase (SOD) is one of the crucial enzymes that protect cells against the oxidative damages (Raychqudhuri & Deng 2000; Fridovich 1986), which is the key enzyme to diminish the concentrations of ROS (Slooten *et al.* 1998). SOD catalyzes the redundant superoxide radicals (O_2^-) to yield molecular oxygen and hydrogen peroxide (H_2O_2). The control of the steady-state O_2^- levels by SOD is an important protective mechanism against cellular oxidative damage, since O_2^- acts as a precursor of more cytotoxic or highly reactive oxygen derivatives, such as peroxynitrite or HO^\cdot (Halliwell & Gutteridge 1999). Therefore, SOD is usually considered the first line of defense against oxidative stress (Sigaud-Kutner 2002). Increased SOD activity was correlated with increased protection from damage associated with oxidative stress (Asada 1999).

Ulmus pumila L. (Ulmaceae), a widespread tree species in North China, with good performance of fast growth, high quality, strong adaptability and resistance, and high economy value, has been used as a major forestation species of shelter forest and saline alkali land in recent years, especially in the semi-arid sand land (Shi 2004; Yan *et al.* 1997). In recent years, some studies have been conducted on this species, but these studies mostly focused on the provenance trails (Wu *et al.* 2001), heredity improvement and variation (Gu *et al.* 1987; Ma *et al.* 1990; Song *et al.* 1995; Sun *et al.* 1999), gas exchange, biomass allocation, preventions of insect pest and disease (Solla *et al.* 2005; Wang and Wang 2005; Qu *et al.* 1999), raising seedling techniques (Zhang *et al.* 2003) of *U. pumila*, and so on. However, little attention has been paid to the permeability and relative antioxidant enzymes activities of protection system in cell plasma membrane for *U. pumila* exposed to salt stress. The roles of SOD under environmental stresses have been studied extensively (Raychqudhuri & Deng 2000; Yu & Rengel 1999), but little is known about the change of SOD activities in *U. pumila* under salt stress. Therefore, in order to clarify the tolerant mechanism of antioxidant enzymes against salt stress, we describe the osmo-

Foundation item: This work was supported by a grant from Heilongjiang Province Science Fund for Distinguished Young Scholars.

Biography: SONG Fu-nan (1972-), male, Ph.D. candidate of Northeast Forestry University, Harbin 150040, P. R. China. Email: sfn@nefu.edu.cn.

Received date: 2005-11-08

Accepted date: 2006-01-12

Responsible editor: Chai Ruihai

*Corresponding author: yangcp@nefu.edu.cn

sis of plasma membrane system and activities of SOD in *U. pumila* leaves exposed to salt stress in this study. This work will provide help for better understanding the roles of SOD in adaptive responses of plant cells under environmental stresses.

Materials and methods

Plant material and stress conditions

One-year-old *U. pumila* seedlings were provided by Fengle Nursery of Zhaozhou County, Heilongjiang Province of China. The samples were transplanted into pots containing riversand pretreated by tap water in greenhouse of Forest Tree Breeding Base of Northeast Forestry University. The seedlings were fertilized complex fertilizer and watered termly to ensure the natural growth of seedlings before salt treatments. Subsequently, the seedlings were treated for seven days by two alkaline salts (NaHCO_3 and Na_2CO_3) selected based on the salt components in the extant salt-alkaline soil of northeast China (Shi and Wang 2005). The mixture of Na_2CO_3 and NaHCO_3 had nine concentrations of 0.3%, 0.6%, 0.9%, 1.2%, 1.5%, 1.8%, 2.1%, 2.4%, 3.0%, and the control. All of treatments were repeated at least five times.

Determination of membrane permeability

Membrane permeability can be reflected by the rate of electrolyte leakage (REL). In our study it was determined as described by Lutts *et al.* (1996). One gram of fresh living leaf was taken from each pot and cut into 1-cm long segments, then washed three times with deionized water to remove surface-adhered electrolytes. The leaf segments were divided equally and placed into two closed vials, each containing 20 mL of deionized water. One vial was incubated at 25°C on a rotary shaker for 3 h, and then the electrical conductivity of the solution (EC1) was determined with a conductivity gauge. The other vial was autoclaved at 120°C for 20 min and electrical conductivity of the solution (EC2) was determined after equilibration to 25°C. REL can be defined as follows:

$$\text{REL (\%)} = (\text{EC1/EC2}) \times 100.$$

SOD activity assay

Ten grams of frozen leaf samples was homogenized on ice with a mortar and pestle for 2 min in 10 mL of homogenizing solution containing 50 mmol·L⁻¹ HEPES buffer and 0.1 mmol·L⁻¹ Na₂EDTA (pH 7.6). The homogenate was centrifuged at 10 000 r·min⁻¹ for 20 min at 4 °C and the supernatant was used for SOD assays (Yu and Rengel 1999).

SOD activity was assayed by monitoring the inhibition of photochemical reduction of nitro blue tetrazolium (NBT), according to the method of Giannopolitis and Ries (1977a, b) with some modifications. For the total SOD assay, a 5-mL reaction mixture, which contains 50 mm HEPES (pH 7.6), 0.1 mmol·L⁻¹ EDTA, 50 mmol·L⁻¹ Na₂CO₂ (pH 10.4), 13 mmol·L⁻¹ methionine, 0.025% (w/v) Triton X-100, 75 μmol·L⁻¹ NBT, 2 μmol·L⁻¹ riboflavin and an appropriate aliquot of enzyme extract, was illuminated for 10 min at a light intensity of 350 μmol·m⁻²·s⁻¹. A control reaction was always performed wherein all the steps and components were exactly the same as described above, except that crude enzyme was replaced with an equal volume of phosphate buffer (pH 7.8) (Sahoo *et al.* 2001). One unit of SOD activity was defined as the amount of enzyme required to cause 50% inhibition of the reduction of NBT as monitored at 560 nm.

Results and analysis

Change of plasma membrane under salt stress

The permeability of plasma membrane was one of the major physiological indexes of osmosis-resistant stress in plants, which reflected directly the stabilization ability of cell for its internal environment and the adaptive and resistant abilities for external environmental change. The effects of salt on structure and constitute of plasma membrane were important aspects of salt stress for plant toxicity, and under high concentration of salt stress, membrane systems of plants were exposed to damage firstly (Zhang *et al.* 2004). With the increase of external salt concentration, the function of cell membrane may be changed by salt ion stress, and intracellular electrolyte began to leak. From Fig. 1, we can see that the rate of electrolyte leakage (REL) changes of *U. pumila* leaves treated presented an increased trend with the increase of salt concentration. At the treatments of salt concentrations less than 1.5%, the REL of samples increased slowly. This phenomenon indicated that cell plasma membrane of *U. pumila* leaves had rather strong resistance to the toxicities of salt ions, and the occurrence of injuries was relative slow. Thus, we can conclude that there exists a half lethal concentration for the injury of cell plasma membrane at the salt concentration less than 1.5%. Whereas, at the salt concentration more than 1.5%, the REL of samples sharply increased, and intracellular electrolyte began to largely leak, which revealed that cell plasma membrane of sample treated had suffered from the severe injury.

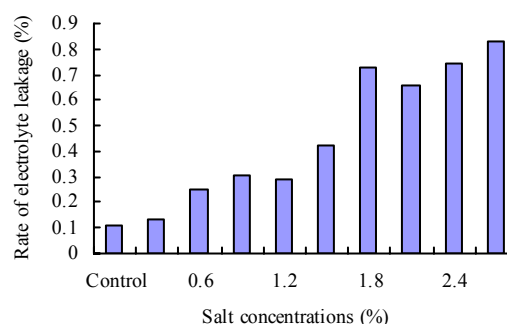


Fig.1 The rate of electrolyte leakage of *U. pumila* leaves at the different salt concentrations

Effect of salt stress on activity of SOD

The activities of SOD were determined in *U. pumila* leaves treated by nine concentrations of salt mixtures. The results showed that SOD of *U. pumila* leaves was induced at salt stress, and the activities of SOD had obvious change (Fig. 2). At the treatments of salt concentrations less than 1.5%, SOD activities presented an increased trend, the growth of seedlings did not decline, and tress and leaves had no symptom of injury. These phenomena showed that the antioxidative defense system in *U. pumila* leaves was not damaged rapidly and can be repaired in a short time under low salt concentrations. While the salt concentrations exceeded 1.5%, SOD activities sharply decreased and REL increased promptly. This indicated that the accumulation of ROS in *U. pumila* leaves had exceeded the regulating thresholds by itself under these conditions. The overproduction of ROS in plant can not be scavenged, which led to decrease or decomposition of SOD activities. Thus, further analyses showed that al-

though plants may enhance their tolerances for salt stress by increasing the activities of SOD to a certain extent, the ROS may still cause the oxidative damage to plasma membrane and inhibit its plant growth when SOD in plants can not scavenge all ROS generated by salt stress.

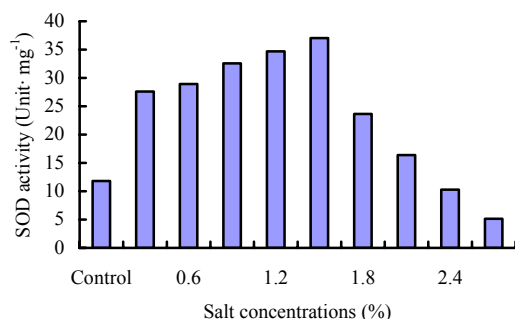


Fig.2 Activities of SOD in *U. pumila* leaves at the different salt concentrations

Discussion

The injuries of osmosis stresses for plants, such as drought, salinity, and chilling, etc., has close relation with oxidative stresses of cell level, and the oxidative stress resistance is also an important measure of salt tolerance for plants. Increased permeability of plasma membrane under salt stress is an expression of plant organ suffering from toxicity (Hasegawa *et al.* 2000). Our study result showed that at the treatments of salt concentrations more than 0.6%, the REL of *U. pumila* leaves also had an increase compared with that of the control. Lower salt concentrations had a slight injury for plant cells, and with salt concentrations increasing, the structure and function of cell plasma membrane were damaged, moreover, various intermetabolic pathways of plants were inhibited, and growth clearly declines (Zhang *et al.* 2003; Xia *et al.* 2005).

The overproduction of intracellular reactive oxygen species (ROS) may result in peroxidation of plasma membrane and oxidative damage (Lin 2004). As a mainly protective mechanism of eliminating ROS in organism, antioxidant enzyme system may enhance the biosynthesis abilities of antioxidant enzymes with the increase of ROS. Thus, the changes of activities of antioxidant enzymes reflects indirectly the existence of harmful substances in circumstance, and it is one of sensitive biological markers in forecasting the toxicities of environmental stresses for ecosystem at molecular levels (Banerjee *et al.* 1999; Li *et al.* 2002). Much evidence obtained from various plant systems showed that the amounts and activities of enzymes involved in scavenging ROS were changed by environmental stresses (Madamanchi *et al.* 1994). Leguminous plants exposed to salt stress showed that the ROS, including O_2^- and H_2O_2 , were generated largely. Though O_2^- and H_2O_2 had a little toxicity for plant, they could be transformed to high toxicity of OH^- through Haber-Weiss reaction. This ROS may damage intracellular biomacromolecule, resulting in DNA injury, enzyme deactivation, and peroxidation of membrane lipid, affecting synthesis and stability of protein, and causing function disorder and cell death (Li *et al.* 2002; Bai and Wang 2002; Pan *et al.* 2003). As an important antioxidant enzyme scavenging ROS in vivo, the induction of SOD activity has been reported by many literatures

for plant subjected to environmental stresses (Madamanchi *et al.* 1994; Chen *et al.* 1999; Hermandwz *et al.* 2002). In our experiment, the induction action of low salt concentrations for SOD activities of *U. pumila* leaves was an adaptive reaction of plant for salt stress to enhance the ability of eliminating ROS. While the inhibition of high salt concentrations for SOD activities can be thought that the tolerance of plant for salt stress has exceeded itself adaptability, and the decrease of SOD activity caused by stress may be regarded as an omen of plant toxicosis (Ireneusz *et al.* 2003; Dong *et al.* 2001; Zhang *et al.* 2005). The results of this study indicated that increased SOD activities may enhance the ability of its scavenging ROS for *U. pumila* to salt stress, and the plant may grow normally. The activities of SOD and the resistance of plant had a certain correlation.

Conclusion

The rate of electrolyte leakage (REL) changes in *U. pumila* leaves treated had an increased trend with the increase of salt concentration. At the treatments of low salt concentrations, the REL of samples increased slowly. This phenomenon indicated that cell plasma membrane of *U. pumila* leaves had rather strong resistance to the injury of salt ion, and the injury occurred relatively slowly. Whereas, at the salt concentration more than 1.5%, the REL of samples sharply increased, and intracellular electrolyte began to largely leak, which showed cell plasma membrane of samples treated had suffered from the severe injury.

The amounts and activities of SOD in *U. pumila* leaves had high levels under salt stress. SOD can make a rapid reaction for environmental stresses, with high abilities in scavenging reaction oxygen species and keeping the balance of reactive oxygen metabolism, and can effectively avoid the oxidative damages of reactive oxygen species for plants under a certain concentration of salt stress.

References

- Alscher, R.G., Erturk, N., Heath, L.S. 2002. Role of superoxide dismutases (SODs) in controlling oxidative stress in plants [J]. *J. Exp. Bot.*, **53**: 1331–1341.
- Asada, K. 1999. The water-water cycle in chloroplasts: scavenging of active oxygen and dissipation of excess photons [J]. *Annu. Rev. Plant Physiol. Plant Mol. Biol.*, **50**: 601–639.
- Bai Hua, Wang Yugu. 2002. Effects of exogenous proline on SOD and POD of activities for soybean callus under salt stress [J]. *Acta Agriculturae Boreali-Sinica*, **17**(3): 37–40. (in Chinese)
- Banerjee, B.D., Seth, V., Bhattacharya, A. 1999. Biochemical effects of some pesticides on lipid peroxidation and free radical scavengers [J]. *Toxicol Letters*, **107**: 33–47.
- Bowler, C., Van Montagu, M., Inzé, D. 1992. Superoxide dismutases and stress tolerance [J]. *Annu. Rev. Plant Physiol. Plant Mol. Biol.*, **43**: 83–116.
- Breusegem, F.V., Vranová, E., Dat, J.E., Inzé, D. 2001. The role of active oxygen species in plant signal transduction [J]. *Plant Science*, **161**: 405–414.
- Chen Wenli, Xu Langlai, Shen Wenbiao and Liu Youliang. 1999. Changes of hydrogen peroxide accumulation and hydrogen peroxide-scavenging enzyme activity in the leaves of two barley species under salt stress [J]. *Journal of Nanjing Agricultural University*, **22**(2): 97–100. (in Chinese)
- Dong Hee Lee, Young Sang Kim, Chin Bum Lee. 2001. The inductive responses of the antioxidant enzymes by salt stress in the rice (*Oryza sativa* L.) [J]. *Journal of Plant Physiology*, **158**: 737–745.

- Fridovich, I. 1986. Superoxide dismutases [J]. *Adv. Enzymol. Relat. Areas Mol. Biol.*, **58**: 61–79.
- Giannopolitis, C.N., Ries, S.K. 1977a. Superoxide dismutase I. Occurrence in higher plants [J]. *Plant Physiol.*, **59**: 309–314.
- Giannopolitis, C.N., Ries, S.K. 1977b. Superoxide dismutase II. Purification and quantitative relationship with water-soluble protein in seedlings [J]. *Plant Physiol.*, **59**: 315–318.
- Greenway, H., Munne, R. 1980. Mechanisms of salt tolerance in non-halophytes [J]. *Ann Rev Plant Physiol*, **31**: 149–190.
- Gu Wanchun, Liu Dean, Tian Yulin. 1987. Selective breeding of *Ulmus pumila* in provenance and family [J]. *Scientia Silvae Sinicae*, **23**(4): 415–424. (in Chinese)
- Halliwell, B., Gutteridge, J.M.C. 1999. Free radicals in biology and medicine, 3rd ed [M]. New York: Oxford University Press.
- Hasegawa, P., Bressan, R.A., Zhu, J.K., *et al.* 2000. Plant cellular and molecular response to high salinity. *Ann Rev Plant Mol Biol*, **51**: 463–499.
- Hermawati, J.A., Almansa, M.S. 2002. Short-term effects of salt stress on antioxidant systems and leaf water relations of pea leaves. *Physiol Plant*, **115**(2): 251–257.
- Ireneusz Ślesak, Zbigniew Misalski. 2003. Superoxide dismutase-like protein from roots of the intermediate C3-CAM plant *Mesembryanthemum crystallinum* L. in vitro culture [J]. *Plant Science*, **164**: 497–505.
- Li Jinting, Zhou Yanqing, Lu Shuxia, Li Ming, Lu Longdou, Ji Xiaoming. 2002. Analysis on the isozymes of superoxide dismutase (SOD) among soybean, cassia obtusifolia and maize seeds [J]. *Journal of Henan Normal University (Natural Science)*, **30**(2): 114–115. (in Chinese)
- Li Weimin, Yin Daqiang, Hu Shuangqing, Chen Liangyan, Wang Liansheng. 2002. Effects of chloric-nitroanilinated chemicals on antioxidant enzymes in serum of *Carassius auratus* [J]. *Acta Scientiae Circumstantiae*, **22**(2): 236–240. (in Chinese)
- Lin Xifeng. 2004. Halophyte research in China [M]. Beijing: Science Press, pp155.
- Lutts, S., Kiner, J.M. and Bouharmont, J. 1996. NaCl-induced senescence in leaves of rice (*Oryza sativa* L.) cultivars differing in salinity resistance [J]. *Ann. Bot.*, **78**: 389–398.
- Ma Changgeng, Li Peiju, Sun Zhansheng. 1990. Ten-year results of filial generation test in *Ulmus pumila* [J]. *Scientia Silvae Sinicae*, **26**(6): 500–505. (in Chinese)
- Madamanchi, N.R., Donahue, J.L., *et al.* 1994. Differential response of Cu, Zn superoxide dismutases in two pea cultivars during a short-term exposure to sulfur dioxide [J]. *Plant Mol Biol*, **26**(1): 95–103.
- Mittler, R. 2002. Oxidative stress, antioxidants and stress tolerance [J]. *Trends Plant Science*, **7**(9): 405–410.
- Pan Dengkui, Zhang Jintong, Yang Jiapou. 2003. Influence of LaCl₃ on activity of Cu, Zn-SOD in soybean [J]. *Journal of the Chinese Rare Earth Society*, **21**(3): 339–342. (in Chinese)
- Qu Qiuyun, Jia Yanmei, Guo Zhonghua, Zhang Jiping, Ye Yongcheng. 1999. A study on occurrence regularity of *Ulmus pumila* withered disease [J]. *Journal of Northwest Forestry University*, **14**(2): 45–50. (in Chinese)
- Raychoudhuri, S.S., Deng, X.W. 2000. The role of superoxide dismutase in combating oxidative stress in higher plants [J]. *Bot. Rev.*, **66**: 89–98.
- Sahoo, R. Kumar, S., Ahuja, P.S. 2001. Induction of a new isozyme of superoxide dismutase at low temperature in *Potentilla astrisanguinea* Lodd. Variety *argrophylla* (Wall. ex. Lehm) Griens [J]. *J. Plant Physiol.*, **158**: 1093–1097.
- Shi Decheng and Wang Deli. 2005. Effects of various salt-alkaline mixed stresses on *Aneurolepidium chinense* (Trin.) Kitag [J]. *Plant and Soil*, **271**: 15–26.
- Shi, L., Zhang, Z.J., Zhang, C.Y., and Zhang, J.Z. 2004. Effects of sand burial on survival, growth, gas exchange and biomass allocation of *Ulmus pumila* seedlings in the Hunshandak Sandland, China [J]. *Annals Botany*, **94**: 553–560.
- Sigaud-Kutner, T.C.S., Pinto, E., Okamoto, O.K., Latorre, L.R., Colepicolo, P. 2002. Changes in superoxide dismutase activity and photosynthetic pigment content during growth of marine phytoplankters in batch-cultures [J]. *Physiol. Plant*, **114**: 566–571.
- Slooten, L., Van Montagu, M., Inzé, D. 1998. Manipulation of oxidative stress tolerance in transgenic plants [C]. in: Lindsey, K. (Ed.), *Transgenic plant research*. Amsterdam: Harwood Academic Publishers, pp241–262.
- Solla, A., Martin, J.A., Corral, P., Gil, L. 2005. Seasonal changes in wood formation of *Ulmus pumila* and *U. minor* and its relation with Dutch elm disease [J]. *New Phytol.*, **166**(3): 1025–1034.
- Song Fuxian, Zhang Jianguo, Liu Huizhong, Li Feng. 1995. Studies on heredity improvement of the second generation of *Ulmus pumila* and its selective breeding of new clones [J]. *Shandong Forestry Science and Technology*, (4): 6–10. (in Chinese)
- Sun Minggao, Zhang Jiguo, Liu Huizhong. 1999. Studies on early selection of siberian elm clones [J]. *Journal of Shandong Agricultural University*, **30**(2): 113–120. (in Chinese)
- Wang Fulin and Wang Dianping. 2005. Control technique of insect pest for planting species (elm) [J]. *Forest By-Product and Specialty in China*, **77**(4): 38. (in Chinese)
- Wang Yihua, Ying Yin, Chen Jia, and Wang Xuechen. 2004. Transgenic Arabidopsis overexpressing Mn-SOD enhanced salt-tolerance [J]. *Plant Science*, **167**: 671–677.
- Wu Guangbiao, Meng Ming, Zhang Fuzhi. 2001. Studies on the provenance trail of *Ulmus pumila* [J]. *J. Shanxi Agric. Uni.*, **21**(3): 285–287. (in Chinese)
- Xia Yang, Liang Huimin, Shu Huairui, Wang Taiming, Liu Dexi, Fang Yifu, Chai Chuanhua. 2005. Changes of leaf membrane penetration, proline and mineral nutrient concentrations of young apple tree under NaCl stress [J]. *Journal of Fruit Science*, **22**(1): 1–5. (in Chinese)
- Xiong, L.M., Schumaker, K.S., Zhu, J.K. 2002. Cell signaling during cold, drought, and salt stress [J]. *Plant Cell*, 165–183.
- Yan Wei, Li Wanbao, Wu Qiaowen. 1997. Lanqi Elms (*Ulmus pumila*) is suitable for plantation in arid grassland [J]. *Journal of Inner Mongolia Forestry College (Nature Science)*, **19**(1): 32–35. (in Chinese)
- Yu, Q. and Rengel, Z. 1999. Drought and salinity differentially influence activities of superoxide dismutases in narrow-leaved lupins [J]. *Plant Science*, **142**: 1–11.
- Yu, Q. and Rengel, Z. 1999. Micronutrient deficiency influences plant growth and activities of superoxide dismutases in narrow-leaved lupins [J]. *Annals of Botany*, **83**: 175–182.
- Zhang Hongbin, Niu Yun, Ge Hongxia, Jia Yuqin. 2003. Introduction of *Ulmus pumila* L. in hexi area of Gansu [J]. *Protection Forest Science and Technology*, **55**(2): 10–12. (in Chinese)
- Zhang Jinfeng, Sun Minggao, Xia Yang, Li Guolei, Hu Xuejian. 2004. Salt stresses affect proline contents, nitrate reductase activities and electrical conductivity of seedling leaves of megranate and cherry [J]. *Journal of Shandong Agricultural University (Natural Science)*, **35**(2): 164–168. (in Chinese)
- Zhang Qiufang, Li Yuanyuan, Pang Caihong, Lu Congming, and Wang baoshan. 2005. NaCl enhances thylakoid-bound SOD activity in the leaves of C₃ halophyte *Suaeda salsa* L. [J]. *Plant Science*, **168**: 423–430.
- Zhang Yubin, Deng Aiyang, Zhuang Tiecheng, Lin Peng. 2003. Relation between soil salinity in intertidal zone and electric conductivity [J]. *Ecology and Environment*, **12**(2): 164–165. (in Chinese)